

Short Sequence-Paper

Full-length cDNA sequences for both ferredoxin-thioredoxin reductase subunits from spinach (*Spinacia oleracea* L.)

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(Received 6 January 1994)

Abstract

Full-length cDNA clones for ferredoxin-thioredoxin reductase subunits A and B of *Spinacia oleracea* were obtained and their complete nucleotide sequences were determined. The results are compared with other known FTR sequences.

Key words: Ferredoxin-thioredoxin reductase; Subunit A; Subunit B; cDNA sequence; (*S. oleracea*)

Ferredoxin-thioredoxin reductase (FTR) is an iron-sulfur protein present in organisms performing oxygenic photosynthesis. It is part of the light-dependent ferredoxin-thioredoxin system which regulates the activity of various chloroplast enzymes by covalent redox modification [1]. FTR has been isolated and characterized in several laboratories [2–4]. The enzyme has an estimated molecular mass of 30 kDa and is composed of two nonidentical subunits, one of which is similar in all organisms studied so far (subunit B: M_r 13 000), while the other is variable in size and characteristic of a particular source (subunit A: M_r 7000 to 16 000) [3]. Both nuclear encoded FTR subunits are thought to be synthesized in the cytosol as precursor proteins containing transit-peptides which are post-translationally processed upon import into the chloroplast. To date, the sequence of the gene encoding the variable subunit A from eukaryotic organisms is still unknown. Only the corresponding gene from the cyanobacterium *Anacystis nidulans* has been cloned and sequenced [5]. The DNA sequence for subunit B from corn and spinach has been reported recently [6].

Using a spinach cDNA library we isolated two full-length cDNA clones which are similar and both encode

subunit A. Moreover, we also isolated a full-length cDNA clone encoding subunit B which is different from the two recently reported sequences from spinach and from corn [6].

For each subunit two degenerate oligonucleotide primers were synthesized based on known amino acid sequences [7].

Subunit A-specific primers:

PFL014, 32-mer sense primer based on 'KQYVGFW-KGKY' (central part):

5'- AA(A/G) CA(A/G) TA(C/T) GTN GGN TT(C/T) TGG AA(A/G) GGN TA -3'

PFL023, 30-mer anti-sense primer based on 'KEEE-FEIIAE' (C-terminus):

5'- (T/C)TC NGC (T/G/A)AT (T/G/A)AT (C/T) TC (A/G)AA (C/T)TC (C/T)TC (C/T)TC (C/T)TT -3'

Subunit B-specific primers:

PFL015, 32-mer sense primer based on 'KDTYFCVD-KCV' (N-terminus):

5'- AA(A/G) GA(C/T) ACN TA(C/T) TT(C/T) TG (C/T) GTN GA(C/T) AA(A/G) TG(C/T) GT -3'

PFL024, 30-mer anti-sense primer based on 'DE-IREVTSNM' (C-terminus):

5'- CAT (A/G)TT N(G/C)(A/T) NGT NAC (C/T)TC NC(T/G) (A/G/T)AT (T/C)TC (A/G)TC -3'

An aliquot (10 μ l) of the spinach cDNA library constructed in λ ZAP II (Stratagene, Heidelberg, Germany) was used directly in PCR reactions with both sets of specific primers [8]. The amplified products

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The sequence data reported in this paper have been submitted to the EMBL Data Library under the accession numbers X77162 (FTR A1), X77163 (FTR A2), X77164 (FTR B).

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Spinach Al    EVALKSDSSTGDFSSSSSSPPPEDEELKKNLEKVGCKVKVKSPLKVYHVP 50
              ||  ||  ||  ||  ||
Anacystis    -----MNVGDRVRVKESVVVYHHP 19

Spinach Al    KLPEVELTP-DMGVIKQYVGFWKGYISPYPFKVEYRIDVPDRGSVKL 99
              |  |  |  |  |  |  |  |  |  |  |  |  |  |
Anacystis    DHRNQAFDLKDAEGEIAAILTEWNGKPISANFPYLVSF-----SNKF 61

Spinach Al    VVHLKEEFEEIIAE 113
              |  |  |  |
Anacystis    LAHLRDFELEVI-- 73

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5'-deletions in order to sequence the unusually long 3'-untranslated trailer regions.

A1	A2	-----CTCATCTCCATCTCAATTATTTCTCATCTAAACAAATCTAAATCTAAACCTCTCTCG	--	-61
A1	A2	AAAAATC		-1
A1	A2	ATGACACACAGGTGTGGCAGTAATGTCTACAGCAACAGCAGCATCAACCGCAACCGCAACG		60
A1	A2	M T T G V A V M S S A T A A S T A T A T		60
A1	A2	GGCGGGCGACGGCGAGATACCACTCTTCCTTAAGCGCAACAACTCATCGGCCACCGTC		120
A1	A2	A A A T A R I P L F L S R N N S S A T V		120
A1	A2	TGCAGCACCTGTAGGTGCAGAACATAACACGAACAAGAACAGAGAGAGCTAGCAATC		180
A1	A2	C S T L R C R T I T R T R T R C G E R L A I		180
A1	A2	TGCTGTGAAGTAGCTTTGAAATCGGATTTCTCAACCGGGTTGGATTTCATCATCATCA		240
A1	A2	C C E V A L K S D S S T G F D S S S S S		240
A1	A2	TCACCTCCAGAAAGACAGGAAATTGAAGAAGAAATCTGGAGAAAGTTGGATTGCAAGGTT		300
A1	A2	S P P E E D E E L K K N L E K V G C K V		297
A1	A2	AAGGTGAATCCCTCTTAAAGTCTACCATGTCTCTAAATTTACCTGAAGTTGAATTAACC		360
A1	A2	K V K S P L K V Y V H V P K L P E V E L T		357
A1	A2	CCAGATATGGTTGGGGTTATTAAAGCAGTAGTTGGATTCTGAAAGGGAAATACATTTC		420
A1	A2	P D M V G V I K Q Y V F W K G K Y I S		417
A1	A2	CCTAATTATCTTTCAAAGTTTGAGTAGCCGATCGATGTCCCTGACCGTGGTTCTGTCAAA		480
A1	A2	P N Y P F K V E Y R I T P D R G S V K		477
A1	A2	TTAGTTGTTCATCTTAAAGAAAGAAGAAATCTGAAATCATAGCAGAGTAGAAGTCTAAATTAT		540
A1	A2	L V V H L K E E E F E I I A E *		537
A1	A2	TTTCTATTTTTATGTGATTGATTATGTATATGTCTTTCTCTTCCCTGTGTGTCTATAC		600
A1	A2	ATTACTACTGATTACTATGCGAGATTGGCAGATGTATCATAAATTTGGCCTTTGTCCAACC		660
A1	A2	CTTAACTTGTCTCTTACTCAATCAGCCCAAGGAGAACATAATGTCTATTTCGCCCTTGC		720
A1	A2	AGT AGT GAAGAGATTAATTAAGAAGTG AA TG TGA GAAGTTAGTAGTAGT AT		717
A1	A2	TGTTGGTGCGACGGCCGAGTTTCGAGTAGTCACAGCGGCACAGCCAAAGGCGCGAGAA		780
A1	A2	ACGATCA TCCCTT AT GCTACT CAATGGCAA TGGTGAAGTT AT T TAT G		777
A1	A2	ACCAAGCAAGAAAGAGCTTTGTAGACGGCTTCTGTGGCAAAATTAACAAAGGAAGTCAAT		840
A1	A2	GATGC C C G G V AT GCTGTAG AA TA GG TATTGTGCTTGGACCG CAT		837
A1	A2	TCATTAAGAACTACCGGATATCTTGGGGGTGGTAGTGTCAAGTGCAGCCGCTCACCAAGT		900
A1	A2	GCGCG TGGCGAG A CAAGAA CA CTCACTGCAA C T GAT T T TGA ACT		897
A1	A2	GAAAGAAAGGGCGACCAACAGTGGAAAGAAAGGACAGATTTCAATTTCCGAGCAGAAGA		960
A1	A2	C C ATGTATA TTA C T T CTG GCT GTGCCCC T ATCAAGT T T		957
A1	A2	AGAACGGTAATGGAGGCTCTTTGGAGGCTCTTTGGCAAGGACTAGTTGATCATAC		1020
A1	A2	TAGT A T AT GCA AG ACCTAA TGG A T T CT TT T C T AT ATTAT		1017
A1	A2	ACCACACTGGTGTAGATGATGATTTCTTCAGTTTCTCGGAGTGTCTTTTGTGCTCTCT		1080
A1	A2	TTAGCTACTACTCTCTGTG ATCCAGTA TA AATAATATCC A A G T T		1077
A1	A2	CTCTCTATATAATAAT		1096
A1	A2	T A G TCT C TT AAATTAAGGATTTGTTTTTAATCAAAACAGTCAATTAATAGTTT		1137
A1	A2	CATCCATGTACTTATTTGTGTATGTACGCTTCTTAGTACTGTGATAGTTTATATAGA		1197
A1	A2	AGTCCCAAAAAATGTTGTTGTG		1220

[illegible]

Fig. 3. Nucleotide sequence and deduced amino acid sequence of a cDNA clone coding for subunit B of ferredoxin-thioredoxin reductase from spinach. The presumed start of the mature protein (i.e., the putative signal sequence processing site) is indicated by a vertical arrow [7.9].

Spinach B1	VEPS-DKSVEIMRKFSQYARKSGTYFCVDKGVTSVVIKGLAEHKDSLGA	49
Spinach B2	ADPS-DKSMEVMRKFSQYFCRKSDTYFCVDKSVTAUVIKGLADHRDTLGA	49
Corn	ADASNDKSVEVMRKFSQYARRSNTFFCADKTVTAUVIKGLADHRDTLGA	50
Spinach B1	PLCPCRYDDKAAEATQGFWNCPCVPMRERKECHCMLFLTPENDFAGKDQ	99
Spinach B2	PLCPCRHYDDKEAEAKQGFWNCPCVPMRERKECHCMLFLTPDNDFAGKEQ	99
Corn	PLCPCRHYDDKAAEVAQGFWNCPCVPMRERKECHCMLFLTPDNDFAGKDQ	100
Spinach B1	TIGLDEIREVTANM	113
Spinach B2	TITLDEIREVTSNM	113
Corn	VISFEEIKEATSKF	114

Fig. 4. Amino acid sequence comparison of various mature ferredoxin-thioredoxin reductase B subunits showing two different spinach polypeptide sequences B1 (this paper), B2 [6] and one from corn [6]. Cysteine residues are shown in bold letters.

respective untranslated regions, particularly in their unusually long 3' ends, this suggests that A1 and A2 represent two different genes. The deduced amino acid sequence of subunit A1 is identical to the published spinach protein sequence [7] except for one amino acid exchange at position 39 where Phe is replaced by Lys in the mature protein sequence (Fig. 2). The coding parts of the subunit A clones exhibit only 25% similarity with subunit A from *Anacystis* and are longer by 40 (39) amino acids (Fig. 2). This confirms the earlier finding that the A subunits from different sources differ significantly in size [3].

In Fig. 3 the nucleotide sequence and the deduced amino acid sequence for a clone encoding FTR subunit B from spinach is shown. The entire coding part comprises 148 amino acids including the plastidic signal sequence. By comparison with the N-terminal amino acid sequence of the protein purified from spinach [7,9] as shown in Fig. 4, the deduced amino acid sequence of its cDNA yields a mature protein of 113 residues (M_r 12 740). There are differences at various positions compared to the previously published sequences from corn and from spinach [6], however, most changes are conservative (Fig. 4). Seven conserved cysteine residues

are present in all three subunit B protein sequences. The additional cysteine residue found in the N-terminal region of the spinach protein [6] is replaced by an alanine residue, as is the case for the corn sequence [6] (Fig. 4).

In contrast to the finding that the FTR subunits A and B differ in size [4] and assuming that the N-termini of both mature subunits [7,9] are complete, our results indicate that both FTR subunits from spinach must be similar in length (113 amino acids). The reason for this discrepancy could be a different processing event in the N-terminus of subunit A in vivo. A further possibility would be an abnormal migration behaviour during SDS-PAGE, as has already been observed for ferredoxin, another small iron-sulfur protein [10].

We are grateful to Dr. U. Sonnewald (Gatersleben) for providing the spinach λ ZAP II cDNA-library and to Dr. H. Lill (Osnabrück) for oligonucleotide synthesis.

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